Understanding the Transmission Dynamics of Anaplasmosis in Cattle

Muhammad Huzaifa¹, Abdullah Siddiqui¹, Fazal Abbas Khan¹, Muhammad Abdul Basit Abbas¹*

1. Riphah international university, Lahore, Pakistan.

*Corresponding author: abdulbasitabbas450@gmail.com

ABSTRACT

Anaplasmosis, an extensive tick-borne infection, emerges from the rickettsial agent Anaplasma marginale, causing increased threats in subtropical and tropical areas. This disease also has zoonotic effects on humans which are very concerning. Female cattle, susceptible due to hormonal fluctuations, milk production, and breeding intricacies, confront increased infection risks, compromising immune resilience. Deep comprehension of transmission modes including biological, mechanical, and transplacental transmission is pivotal for effective control strategies, grappling with the complexity of global vector dynamics and the multifaceted nature of anaplasmosis

Introduction:

Anaplasmosis in cattle results from the presence of Anaplasma (A.) marginale, a rickettsial organism. It is the most prevalent tick-borne disease and is particularly dominant in cattle living in subtropical and tropical regions, where it can cause significant morbidity and mortality in susceptible animals (1). The geographic distribution of this disease depends upon the presence of its vector along with its reservoir host (2). This disease is also zoonotic so we should deal with this disease carefully. This infection affects females more than males (3). There are three types of transmissions.

Biological Transmission

In cattle Ixodidae family (hard tick) of the ticks transmits A. marginale. Majorly involved genera are Riphicephalus, Dermacentor, Ixodes, and Hyalomma. A. marginale replicates in the midgut of the tick from where it moves to the salivary gland. When the tick sucks blood from the host it leaves anaplasma in the blood of the host. Riphicephalus species are major vectors of A. marginale in Australia and Africa. Their prevalence in Pakistan is low as compared to other ticks. Among all the species Riphicephalus (R.) microplus is one of the major species that transfer the anaplasmosis. R. microplus is a one-host tick which means it spends its whole life on a singlehost species. Because of the migratory role of the male ticks they transmit disease from infected to healthy animals more often. Anaplasma shows transtadial and intrastadial transmission in ticks. It is also seen that A. marginale shows transovarial transmission in R. microplus (4). Dermacentor species are major vectors of A. marginale in USA. Dermacentor is a threehost tick. A. marginale transmits transcardially but does not show transovarian route of transmission. Dermacentor (D.) andersoni female get infected at the nymph stage and transmit the infection on the 6th to 7th day of feeding on the host. On the other hand, males transmit the infection within 24 hours (5). Ixodes are present all around the world but found particularly in the Northern and Eastern Midwest of the USA and in Southeastern Canada. It is also a three-host tick. Transmission through these ticks is less as compared to Riphicepalous and dermacentor. A marginale is transmitted transcardially in ixodes. The ticks of Hyalomma genus are present near Pak-Iran border and their surrounding areas and they have the highest prevalence for transmitting A. marginale in cattle in Pakistan.

Mechanical Transmission

Transmission through Blood Sucking Flies

Blood-sucking flies in the family *Tabanidae* and *Muscidae* act as a vector for the transmission of anaplasmosis. Mechanical transmission of A. marginale by Horse fly and Stable fly is seen in the USA. When they suck the blood from infected animals their external mouthparts get contaminated with infected blood and when they move to another host the contaminated blood from their mouthparts moves into the body of the host. Horse flies can transfer the disease for up to two hours after feeding on an infected animal (7). In areas where ticks are lesser in number, like in Uruguay, horseflies could be the important vector for A. marginale. So for the confirmation, in Uruguay, eight horsefly species were tested (Dasybasis missionum, Poeciloderas lindneri, Tabanus campestris, T. claripennis, T. fuscofasciatus, T. platensis, T. tacuaremboensis and T. triangulum). Four species were found to be positive for A. marginale, with D. missionum and P. lindneri having the highest number of infections, while only one individual each of T. fuscofasciatus and T. tacuaremboensis was positive. Both D. missionum and P. lindneri were positive for A. marginale in areas where ticks are less (6). Stable flies can transmit that particular strain of A. marginale in cattle. A study of dairy cows in Costa Rica shows that Tabanidae and stable fly are important in the transfer of A. marginale (13).

Transmission through blood contaminated equipment's

Published on: 1 December 2023

Equipment's like vaccination needles which plays important role in the transmission of the pathogen. Other examples include the use of dehorner, and other surgical instruments. Transferring blood from an infected or reservoir animal can cause anaplasmosis (7).

Transplacental Transmission

This type of transmission occurs when the pathogen is transmitted from dam to fetus. It occurs in the second and third trimester of the pregnancy. A study in Brazil shows that 10 percent of calves born to infected mother were infected with anaplasmosis (8). According to another study 16 percent of calves were born infected (9).

Conclusion

In conclusion, to control anaplasmosis (the most prevalent tick-borne disease) we have to overcome the routes of its transmission. To this end, we must use sterilized surgical instruments and apply acaricides. Other than that, management practices at farms must be improved for example closure of cracks and crevices. To control flies, we should keep the nearby places clean so that larvae of flies cannot get a suitable environment for their growth

References

- Haji I, Simuunza M, Kerario II, Jiang N, Chen Q. Epidemiology of tick-borne pathogens of cattle and tick control practices among mixed farming and pastoral communities in Gairo and Monduli districts, Tanzania. Veterinary Parasitology: Regional Studies and Reports. 2022 Jul
- Singh H, Haque M, Singh NK, Rath SS. Molecular detection of Anaplasma marginale infection in carrier cattle. Ticks and tick-borne diseases. 2012 Feb 1;3(1):55-8.

 Sajid MS, Siddique RM, Khan SA, Zafar I, Khan MN. Prevalence and risk factors of
- anaplasmosis in cattle and buffalo populations of district Khanewal, Punjab, Pakistan. Global Veterinaria, 2014:12(1):146-53
- De La Fourniere S, Guillemi EC, Paoletta MS, Pérez A, Obregón D, Cabezas-Cruz A, Sarmiento NF, Farber MD. Transovarial transmission of anaplasma marginale in Rhipicephalus (Boophilus) microplus ticks results in a bottleneck for strain diversity. Pathogens. 2023 Aug 2.12(8).1010
- Stich RW, Kocan KM, Palmer GH, Ewing SA, Hair JA, Barron SJ. Transstadial and attempted transovarial transmission of Anaplasma marginale by Dermacentor variabilis. American journal of veterinary research. 1989 Aug 1;50(8):1377-80.
- of veterinary research. 1893 Aug 1,50(3):1371-60.
 Rodrigues GD, Lucas M, Ortiz HG, dos Santos Gonçalves L, Blodorn E, Domingues WB, Nunes LS, Saravia A, Parodi P, Riet-Correa F, Menchaca A. Molecular of Anaplasma marginale Theiler (Rickettsiales: Anaplasmataceae) in horseflies (Diptera: Tabanidae) in Uruguay. Scientific Reports. 2022 Dec 28;12(1):22460.

 Abdisa T. Epidemiology of Bovine Anaplasmosis. SOJ Vet Sci. 2019;5(1):1-6.
- Grau HE, Cunha Filho NA, Pappen FG, Farias NA. Transplacental transmission of Anaplasma marginale in beef cattle chronically infected in southern Brazil. Revista Brasileira de Parasitologia Veterinária. 2013 Apr;22:189-93.
- Salabarria FF, Pino R. Vertical transmission of Anaplasma marginale in cows affected in late pregnancy. Revista Cubana de Ciencias Veterinarias. 1988;19:179-82.